

## A Study on the Protective Effect of Single Dose of N-2- Mercaptopropionyl Glycine Administration Prior to Renal Reperfusion on Wistar Rats

<sup>1</sup>Vinodini N.A., <sup>2</sup>Tripathi Y., <sup>3</sup>C.V. Raghuveer, <sup>4</sup>Asha Kamath, <sup>5</sup>C. Ramaswam

<sup>1</sup>Department of Physiology Kasturba Medical College Mangalore (Unit of Manipal university  
Manipal Karnataka India)

<sup>2</sup>Tripathi Y Present address: P.O.Box No:64832, Ghariyan, Libya

<sup>3</sup>Department of pathology Malaka Manipal College Manipal Karnataka India

<sup>4</sup>Department of community Medicine Kasturba medical College Manipal Karnataka India

<sup>5</sup>Department of physiology Saveetha Medical College Chennai India

---

**Abstract:** N-2-mercaptopyrionylglycine (NMG) is a powerful antioxidant which inhibits synthesis of superoxide radicals. It has been tested as a preventive agent against metabolic and structural damage induced during ischemia/reperfusion (I/R) process. To validate the efficacy of NMG on renal tissue this work was devised. During the experiment, the wistar strain rats were divided into five groups; the group I as normal control, the group II & III as I/R controls, and the group IV & V as I/R treated groups. The I/R control and I/R treated groups were subjected to ischemia and reperfusion either for 10 minutes (Gr.II & IV) or 90 min. (Gr.III & V). The I/R treated groups in addition received N-2- mercaptopropionylglycine (50mg/kg bw) intravenously prior to the onset of reperfusion. The results suggest that 60 minutes of ischemia followed by 10 (Gr.II) and 90 minutes (Gr.III) of reperfusion increased lipid peroxidation (MDA), and decreased glutathione (GSH), & Superoxide dismutase levels in the renal tissues compared to normal control. However, the GSH and SOD levels were higher and the lipid peroxidation level was lower in I/R treated groups than I/R controls. Thus, the present study confirms the protective effect of N-2- mercaptopropionylglycine on the reperfusion injury and also extends that protective effect on renal tissue.

**Key words:** ischemia, reperfusion, NMG, MDA, GSH, SOD.

---

### INTRODUCTION

Renal ischemia initiates a complex and interrelated sequence of events, resulting in injury and death of the renal cells (Dong Kyun Rah et al ,2007). Reperfusion, although essential for the survival of the ischemic tissue, causes additional damage (termed as reperfusion injury (Lieberthal W and Levine JS, 1996). Together, I/R of the kidney contribute to the renal dysfunction and associated with ischemic acute renal failure (Gueller, F et al, 2004). Interruption of blood circulation produces biochemical and structural cell disintegration that evolve to organelle disorganization with subsequent cellular death (Souza AP et al 1991; & Lefler AM, Lefler DJ, 1993). The oxidative-reduction activity, mainly mediated by newly formed highly reactive chemical radicals, is one of the processes responsible for this condition (Del Maestro RF 1980; & Gao W 1991).

When ischemic tissue is reperfused, high concentrations of xanthine-oxidase will use the incoming oxygen to produce hydro peroxide and subsequently liberate great amounts of aggressive free radicals (Souza AP et al 1991). The dissociation of oxygenated water by metallic ions such as iron and copper stimulates the production of free radical hydroxyls and is highly harmful to the reperfused tissue causing rupture of cellular and sub-cellular structures (Del Maestro RF 1980; Belzer FO, Southard JH.1988). The anti oxidative effect of N<sub>2</sub>-mercaptopyrionylglycine (N<sub>2</sub>-MPG), that partially prevents the conversion of xanthine-dehydrogenase into xanthine-oxidase, was previously demonstrated by the preservation of miocutaneous graft after 12 hours of normothermic ischemia (Emilio Elias Abdo et al 2003). The tissue protective effect of N<sub>2</sub>-MPG was also demonstrated in the acute pancreatitis model, where low blood flow and cellular damage, resembles the ischemia/reperfusion process (Lefler AM, Lefler DJ 1993). But, not a single report was available about protective role of N<sub>2</sub>-NMG on renal tissue in limiting free radical-induced tissue reperfusion damage. Therefore, the present study was designed to find out the importance of the protective effect of N-2- mercaptopropionyl glycine on renal tissue I/R.

---

**Corresponding Author:** Dr Vinodini NA Associate professor Department of physiology center for Basic Sciences  
Kasturba Medical College Bejai Mangalore Karnataka 575004 Mob : 919448214870

## MATERIAL AND METHODS

### **Animals:**

Inbred wistar rats (WI) weighing about 200-300gm were used in this study. The rats were obtained and maintained in the Animal house of the department of physiology center for Basic Sciences Kasturba Medical College Mangalore. All experimental protocols were approved by the ethical committee of Manipal University Manipal, Karnataka. During the experimental periods, the rats were kept in an individual cage and food & water were given ad lib.

### **Grouping Protocol:**

**Group I:** (n=7; Normal control). The animals in this group served as control.

**Group II & III:** (n=7 each; I/R control) In this; the rats underwent ischemia for 60minutes followed by 10 minutes (II) & 90 minutes (III) of reperfusion respectively.

**Group IV & V:** (n=7each; I/R treated group) The rats underwent 60 minutes of ischemia and were treated with a loading dose of NMG (50mg/kg/bwt) prior to the commencement of reperfusion for 10 minutes (IV) & 90 minutes (V) respectively.

### **Procedure for Ischemia/reperfusion:**

The rats were anesthetized intraperitoneally with pentobarbitone sodium (40mg/kg/bw) under strict aseptic conditions. The abdomen was opened by left flank incision. Left renal artery and vein were occluded by micro vessel occluder. The animals were subjected to renal ischemia for 60 minutes. Renal ischemia was confirmed by inspection of the renal vessel. At the end of ischemic period the vessel occluder was removed for reperfusion of the kidney for 10 minutes (II & IV) or 90 minutes (III & V). The Gr.IV & Gr.V also received a loading dose of NMG (50/kg/bwt) prior to the onset of reperfusion. The abdominal viscera were covered with gauze soaked in normal saline (0.9% sodium chloride) to keep the tissues moist.

### **Procedure for Harvesting the Tissue:**

Following completion of ischemia and reperfusion, the kidneys were removed and kept in cold phosphate buffered saline (PBS, 0.9%). The reperfused kidney was blotted, dried and minced. The minced tissues were transferred to a glass homogenizer containing 10 ml of cold PBS (PH 7.4) and were centrifuged at 3000 rpm for 30 minutes. The supernatant obtained was used for the biochemical estimation such as MDA, GSH and SOD.

### **Estimation of MDA:**

MDA was estimated by the method described by Kartha & KrishnaMurthy (1978). The development of pink color was measured at 535nm by using a Spectronic D-20 Spectrometer. TBA- reactive material was expressed in terms of nano moles of malondialdehyde (MDA)/gm of wet tissue.

### **Estimation of GSH:**

GSH was estimated standard protocol (Beutler E, etal, 1963). The OD of the blank test tube with all the reagents except the samples and the test tube with all the reagents with the sample were measured at 412 nm by using a Spectron D-20 spectrometer. The glutathione content of tissue was expressed as GSH in  $\mu\text{g/gm}$  of tissue protein.

### **Estimation of SOD:**

SOD was estimated by the technique explained by Fridovich(1971).The reduction of NBT by  $\text{O}_2^-$  was measured at 560 nm using a Spectron D-20 Spectrophotometer. The activity was expressed as unit/ mg protein

### **Statistical Analysis:**

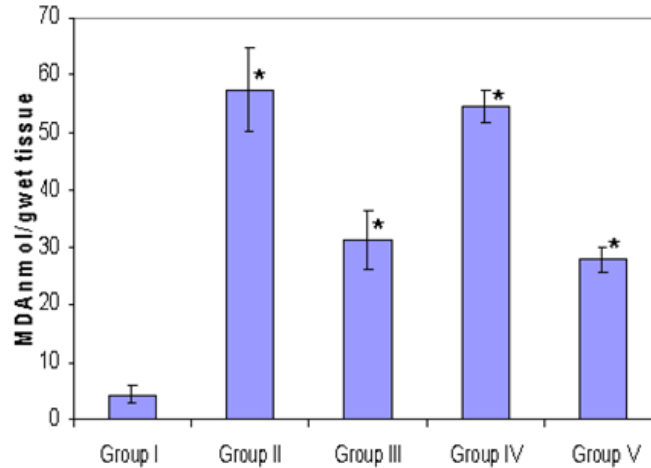
The data were expressed as means  $\pm$ SD from 7 animals per group. The differences between the groups were compared for statistical significance by the student t test with the level of significance set at  $P<0.05$ .

### **Results:**

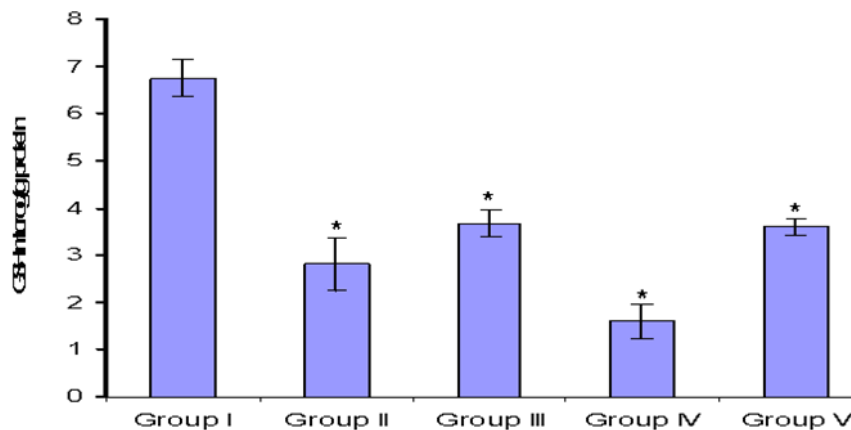
The MDA level, the measure of lipid peroxidation increases significantly in both I/R control groups (Gp II & III) ( $p<0.0001$ ) when compared with normal control (Gp I). However, administration of N-2-

mercaptopyropionyl glycine (NMG) prior to reperfusion decreased the MDA level. As a result, I/R treated groups (Gp. IV & V) showed a decrease in MDA level with both I/R controls (P<0001 for all). (Fig: 1).

The I/R control groups II & III showed a decrease in GSH and SOD level in comparison with the normal control, group I (p<0.0001). But the GSH & SOD level of these I/R control groups (II & III) were significantly more than the I/R treated groups of IV & V (P< 0.0001 for all) (Fig: 2 & Fig;3).



**Fig. 1:** Effect of NMG administration prior to reperfusion on tissue level of MDA following renal ischemia Gr. I vs Gr. II, Gr. III, Gr. IV and Gr. V n-sample size=7 p<0.0001.

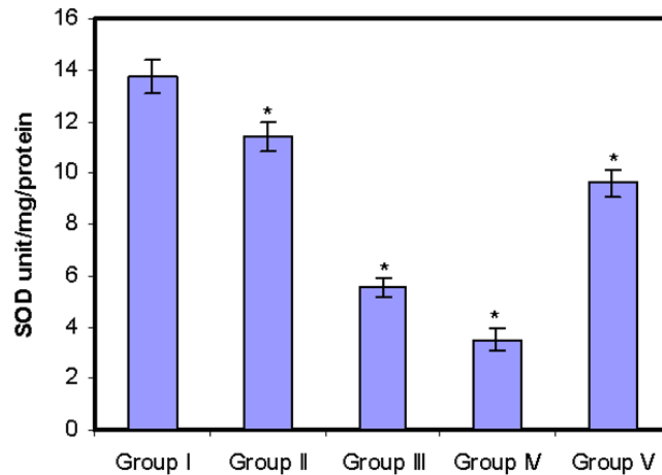


**Fig. 2:** Effect of NMG administration prior to reperfusion on tissue GSH level following renal ischemia Gr. I vs Gr. II, Gr. III, Gr. IV and Gr. V n-sample size=7 p<0.0001.

**Discussion:**

Oxygen free radicals are usually produced by the aerobic metabolism of living cells. An overproduction of these radicals has been reported in ischemic/reperfused organs, leading to a high degree of tissue damage (Epstein FH 1985, Granger DN et al 1989, Kobelt F, et al 1994 Vento AE etal 1999). Prevention of these deleterious effects, by the administration of antioxidative substances has been attempted in organ transplantation and surgical emergencies (.Horwitz LD et.al 1994. Ihnken K et al, 1995, Vento AE etal 1999).

After a significant period of ischemia, blood recirculation produces large amounts of super oxide residues derived from the abnormal activity of the newly formed xanthine-oxidase and is responsible for tissue lesion (Del Maestro RF etal 1980 ,Widman SC et a 1994). The N<sub>2</sub>-MPG prevents the formation of xanthine-oxidase (Emilio Elias Abdo 1988) and thereby reduces the reperfusion injury by decreasing the production of oxidative free radicals. The result of the present study which shows, a decrease in the level of tissue peroxidation and increase in the level of GSH & SOD, after the administration of N<sub>2</sub>-MPG prior to reperfusion following ischemia also in agreement with the above statement. As the protective effect of N<sub>2</sub>-MPG in this study is more evident with group received reperfusion of 10 minutes than the group received 90 minutes reperfusion indicates that the protective action of N<sub>2</sub>-MPG is short lived.



**Fig. 3:** Effect of NMG administration prior to reperfusion on tissue SOD level following renal ischemia Gr. I vs Gr. II, Gr. III, Gr. IV and Gr. V n-sample size=7 p<0.0001.

Thus, the results of this study support the present notion that N<sub>2</sub>-MPG is potent in decreasing the renal injury mediated due to oxidative stress which is responsible for tissue destruction. This study also suggest for to get better protective effect against reperfusion injury at least in renal tissue, along with a single dose of prior administration, repeated shots of N<sub>2</sub>-MPG at certain intervals during reperfusion may be needed; a new finding has to be investigated further.

#### REFERENCES

- Abdo, E.E., S.N. Sampietri, A.M. Coelho, N. Molan, M. Kubrusly, 1998. N<sub>2</sub>-Mercaptopropionylglycine(N<sub>2</sub>-MPG) in experimental acute pancreatitis. *Rev Hosp Clin Fac Med S Paulo*, 53: 169-73.
- Beauchamp, C., I. Fridovich, 1971. Superoxide dismutase. Improved assay and assay applicable to acrylamide gel. *Ann Biochem.*, 44: 276-287.
- Belzer, F.O., J.H. Southard, 1988. Principles of solid-organ preservation by cold storage. *Transplantation*, 45: 670-3.
- Beutler, E., O. Duron, B.M. Kelly, 1963. Improved methods for the determination of glutathione. *J Lab Clin Med.*, 61: 882-888.
- Cai, M., R. Ogawa, 1994. Effect of free radical scavengers, methylprednisolone and ulinastin on acute xanthine and xanthine-oxidase induced lung injury in rats. *Circ Shock*, 43: 71-8.
- Del Maestro, R.F., 1980. An approach to free radicals in medicine and biology *Acta Physiol Scand; Suppl*, 492: 153-68.
- Dong Kyun Rah, Dong Wook Han, Hyun Sook Back, Suong Hyu Hyon, Beyoung Yun Park and Jong Chul Park, 2007. Protection of rabbit kidney from ischemia/ reperfusion injury by green Tea phenol pretreatment. *Arch Pharm Res*, 30(11): 1447-1454.
- Milio Elias Abdo, José Eduardo Monteiro Cunha, Pérsio Deluca, Ana Maria Mendonça Coelho, Telesforo Bacchella, Marcel Cerqueira César Machado, 2003. Protective effect of N<sub>2</sub>-mercaptopyronylglycine on rats and dogs liver during ischemia/reperfusion process. *Arq. Gastroentero*, 40: 3.
- Epstein, F.H., 1985. Oxygen-derived free radicals in post ischemic tissue injury. *N Engl J Med.*, 312: 159-63.
- Gao, W., Y. Takey, I. Marzi, K.A. Lindert, J.L. Caldwell-Kenkel, R.T. Currin, Y. Tanaka, J.J. Lemasters, R.G. Thurman, 1991. Carolina rinse solution - a new strategy to increase survival time after orthotopic liver transplantation in the rat. *Transplantation*, 52: 417-24.
- Granger, D.N., J.N. Benoit, M. Suzuki, M.B. Grisham, 1989. Leukocyte adherence to venular endothelium during ischemia-reperfusion. *Am J Physiol.*, 257: G683-8.
- Gueller, F., W. Gwinner, A. Schwarz and H. Haller, 2004. Long-term effects of acute ischemia and reperfusion injury. *Kidney Int.*, 66: 523-527.

- Horwitz, L.D., P.V. Fenessey, R.H. Shikes, Y. Kong, 1994. Marked reduction in myocardial infarct size due to prolonged infusion of an antioxidant during reperfusion. *Circulation*, 89: 1792-801.
- Ihnken, K., K. Morita, G.D. Buckberg, M.P. Sherman, H.H. Young, 1995. Studies of hypoxemic/reoxygenation injury without aortic clamping. Counteraction of oxidant damage by exogenous antioxidants: N2-mercaptpropionylglycine and catalase. *J Thorac Cardiovasc Surg*, 110: 1212-20.
- Kobelt, F., U. Schreck, H.A. Henrich, 1994. Involvement of liver in the decompensation of hemorrhagic shock. *Shock*, 2: 281-8.
- Kartha, V.N.R., S. Krishnamurthy, 1978. Factors effecting in vitro lipid peroxidation in rat homogenate. *Ind J Physiol Pharmacol.*, 22: 44-52.
- Lefer, A.M., D.J. Lefer, 1993. Pharmacology of the endothelium ischemia-reperfusion and circulatory shock. *Ann Rev Pharmacol Toxicol.*, 33: 71-90.
- W and J.S. Levine, 1996. Mechanisms of apoptosis and its potential role in renal tubular epithelial cell injury. *Am J Physiol.*, 271: F477-F488.
- Lowry, O.H., N.J. Roscbrought, A.I. Faro, R.J. Randal, 1951. Protein measurement with folin phenol reagent. *J Biol Chem.*, 193: 265-275.
- Souza, A.P., A.R.B. Coelho, R.D. Camara-Neto, E.M. Ferraz, J.V. Rodrigues, J.F.C. Lima-Filho, A.M.S. Telles, 1991. Complete acute occlusion of the portal vein in dogs. A study based on hemodynamics and hematologic data aimed at its application in liver transplantation. *ABCD Arq Bras Cir Dig.*, 6: 15-9.
- Vento, A.E., O.J. Rämö, A.T. Nemlander, M. Ahotupa, E. Nissinen, A. Holopainen, S.P. Mattila, 1999. Nitecapone inhibits myeloperoxidase in vitro and enhances functional performance after 8 h of ischemia in experimental heart transplantation. *Res Exp Med (Berl)*, 198: 299-306.
- Widman, S.C., C.S. Liang, E.A. Schenk, W.B. Jr. Hood, 1994. Contraction band necrosis: its modification by the free radical scavenger N2-MPG. *J Cardiovasc Pharmacol.*, 24: 694-701.